• Bloodworks Labs

REQUEST FOR TESTING

LABORATORY SERVICES

Immunohematology Referen 921 Terry Ave. Seattle, WA 98104 Phone 206-689-6534 Fax 206-689-8 Laboratory Staffed for Questions 24/7		oratory (IRL)	Spee	cimen Number	Order Number
For current test descriptions and CPT codes visi	it <u>https://ww</u>	w.bloodworksnw.org/labs/test	<u>s</u>	BW Tech ID	TIME RECEIVED
TESTING PROFILES – May include one or	more of the	e individual tests given below	w.		
 Antibody Identification Hemolysis evaluation Suspected delayed hemolytic transfusior Prenatal Antibody Identification (includes) 		tration, if indicated)	Resolution	n marrow transplant (HSCT) re n of ABO discrepancy utination (includes lectin panel, ease specify)	if indicated)
INDIVIDUAL TESTS – If Profile has been cl	hecked abo	ve, do NOT check test belo	W.		
3103-03 ABO & Rh (D antigen try 3103-03 Solid Organ Donor ABO 3105-00 % ABO for HSCT 3104-02 Indirect Antiglobulin Te 3125-02/01 Direct Antiglobulin Test 3129-00 Elution 3117-00/3118-00 Sickle Cell Phenotype (D, C, E 3136-00 Extended Patient Phenotype 3118-00 Single Antigen Phenotype	D & Rh (A1 st (antibody t (polyspeci (Rh & K) , c and e ar otype (7 or	v screen) fic and monospecific) ntigen typing)	3139-00	•	nsplant Titer (⊡anti-A or ⊡anti-B) splant Titer (⊡anti-A or ⊡anti-B) (anti-A or anti-B) ti-A or anti-B)
GENOMICS TESTS (If genomics testing is 3117-08 ABO Genotyping Reason for DNA Analysis:*Note: For add		please check a box below	3117-04		iple Blood Groups
PLEASE PRINT. Submit separate request and NOTE: Information in RED must be completed.		ood sample per laboratory.	Contact	Person:	
Sample Drawn: DATE /	TIME	am/pm 2 nd person reviewing patient ID If Required by facility policy)	REPOR	Name DE PHONE NUMBER OR FAX N T RESULTS AS SOON AS AVA & STAT TESTING	
Specimen/Accession No.:			If results	are needed as soon as available	
Physician or Authorized Person Order Test Diagnosis/Purpose of Testing:			Nam		() Number
History / Comments / Special Instructions:			Name		
Form Completed By:					
Name must match EXACTLY name on sample Name on Sample LAST F	label. FIRST	MIDDLE	Name		
Hospital Identification Number			City, S	State, Zip	
Hospital/Institution					
Social Security Number	Sex (M/F)	Date of Birth (mm/dd/yr)			

Instructions:

1. Ensure sample is of appropriate type, labeled correctly and the required volume of blood is provided. Do not use tubes that contain a silicone separator gel. Additional information on sample requirements, CPT codes, test description, scheduling and reporting can be found at https://www.bloodworksnw.org/lab/tests

Test	Requested Amount			
Antibody Identification	2 full 7 ml EDTA tubes OR 2 10 ml clotted sample.			
Prenatal Antibody Identification (titration)	*see note below for minimum sample requirements			
ABO antibody Titers				
HSCT Long term Follow-up	2 full 7 ml EDTA tubes			
Suspected Delayed Hemolytic Transfusion Reaction	*see note below for minimum sample requirements			
Hemolysis Evaluation				
NOTE: Minimum sample requirement for above tests: One f	full 7 ml EDTA sample as the minimum amount.			
Patients 1 - 5 years old: One full 3 ml EDTA sample mi	nimum.			
Patients ≤1 year old: Two full 0.5 ml EDTA microtaine	rs (1.0 ml total) of peripheral blood is the minimum amount.			
Donath-Landsteiner Test	10 ml clotted sample drawn and maintained at 37°C until serum is separated from			
Donath-Lanusteiner Test	clot.			
Thermal Amplitude Test	10 ml clotted or 7 ml EDTA sample maintained at 37°C until serum/plasma is			
Thermal Amplitude Test	separated			
ABO Genotyping	7ml EDTA tubes			
Red Cell Genotyning for Multiple Blood Groups				

Red Cell Genotyping for Multiple Blood Groups

Contact IRL for sample requirements for any special testing not listed above.

Sample Labeling: All samples must be properly labeled and information must agree with the identification on the RFT.

- The sample requires two patient unique identifiers. If a sample is identified by name, there must be a numeric identifier which may include hospital number or other coded identifier. A birthdate is not acceptable in this circumstance.
- A draw date should be on the sample.

<u>When RBCs are to be crossmatched</u>: Samples must additionally include: full name of patient, date and time obtained, hospital and/or patient identification number and the identification of the individual obtaining the sample. Submit a Request for Blood and Blood Components form. Also notify the laboratory for additional sample requirements.

2. Complete the IRL Request for Testing form (RFT); it <u>must</u> contain all of the information that is printed in red: draw date/time, physician or authorized person ordering test, to whom to send the report. Identifying a contact person is required to facilitate timely resolution of discrepancies and questions.

3. Complete and send the Immunohematology Consultation Request Form. Include copies of serological evaluations worksheets.

4. Additional samples: If patient has been transfused within the last 30 days, and submitting for antibody identification, send pre- and post-transfusion samples.

5. Notification and Transport: Notify the laboratory of shipping arrangements by phone (206-689-6534). All samples must be sent to Bloodworks Northwest in a sealed, leak-proof container marked with a biohazard sticker to comply with OSHA safety standards. Ship at ambient temperature unless instructed otherwise.

Tests that may be performed in Profiles:

Antibody Identification		Prenatal Antibody Identification	Hemolysis evaluation		
Red cell panel	3126-00	Red cell panel	3126-00	Red cell panel	3126-00
Direct Antiglobulin Test-Polyspecific	3125-02	Direct Antiglobulin Test-Polyspecific	3125-02	Direct Antiglobulin Test-Polyspecific	3125-02
Direct Antiglobulin Test-Monospecific	3125-01	Direct Antiglobulin Test-Monospecific	3125-01	Direct Antiglobulin Test-Monospecific	3125-01
Elution	3129-00	Elution	3129-00	Indirect Antiglobulin Test -Antibody screen	3104-02
Extended Patient Phenotype	3136-00	Extended Patient Phenotype	3136-00	Elution	3129-00
Rh & K Phenotype	3117-00	Rh & K Phenotype	3117-00	Positive antibody screen reflex to Antibody	
	3118-00		3118-00	Identification as indicated	
Red cell separation for phenotyping	3131-00	Red cell separate for phenotyping	3131-00		
Auto/allo adsorption	3127-00	Auto/allo adsorption	3127-00	Polyagglutination	
	3128-00		3128-00		
Chemical treatment of Red Cells	3132-00	Chemical treatment of Red Cells	3132-00	Minor crossmatch with adult and cord sera	3137-01
Enzyme Treatment of Red Cells	3133-00	Enzyme treatment of Red Cells	3133-00		
Chemical Treatment of Serum	3134-00	Chemical Treatment of Serum	3134-00		
Neutralization	3130-00	Neutralization	3130-00	Reflex to lectin panel if indicated	3137-02
Red cell antibody Titrations	3115-00	Red cell antibody Titrations	3115-00		
Note: Reflex to red cell genomics testing if in	dicated	Note: Reflex to red cell genomics testing if indicate	ed		
Suspected delayed hemolytic transfusion reaction		Long term marrow transplant (HSCT) recipie	Resolution of ABO discrepancy		
Direct Antiglobulin Test-Polyspecific	3125-02	Direct Antiglobulin Test-Polyspecific	3125-02	Direct Antiglobulin Test-Polyspecific	3125-02
Direct Antiglobulin Test-Monospecific	3125-01	Direct Antiglobulin Test-Monospecific	3125-01	Direct Antiglobulin Test-Monospecific	3125-01
Elution	3129-00	ABO/Rh typing	3103-03	Extended Patient Phenotype	3136-00
Red Cell Panel	3126-00	%ABO	3105-00	RBC Phenotype- Single antigen	3118-00
AHG crossmatch of implicated RBCs unit	3015-02	Indirect Antiglobulin Test-antibody screen	3104-02	Chemical treatment of Red Cells	3132-00
Positive AHG Crossmatch reflex to		Anti-A/Anti-B Titrations	3115-00	Chemical treatment of Serum	3134-00
Antibody Identification as indicated		Red cell panel	3126-00	Red cell panel	3126-00
		Positive antibody screen reflex to		Rh & K Phenotype	3117-00
		Antibody Identification as indicated			3118-00